

---

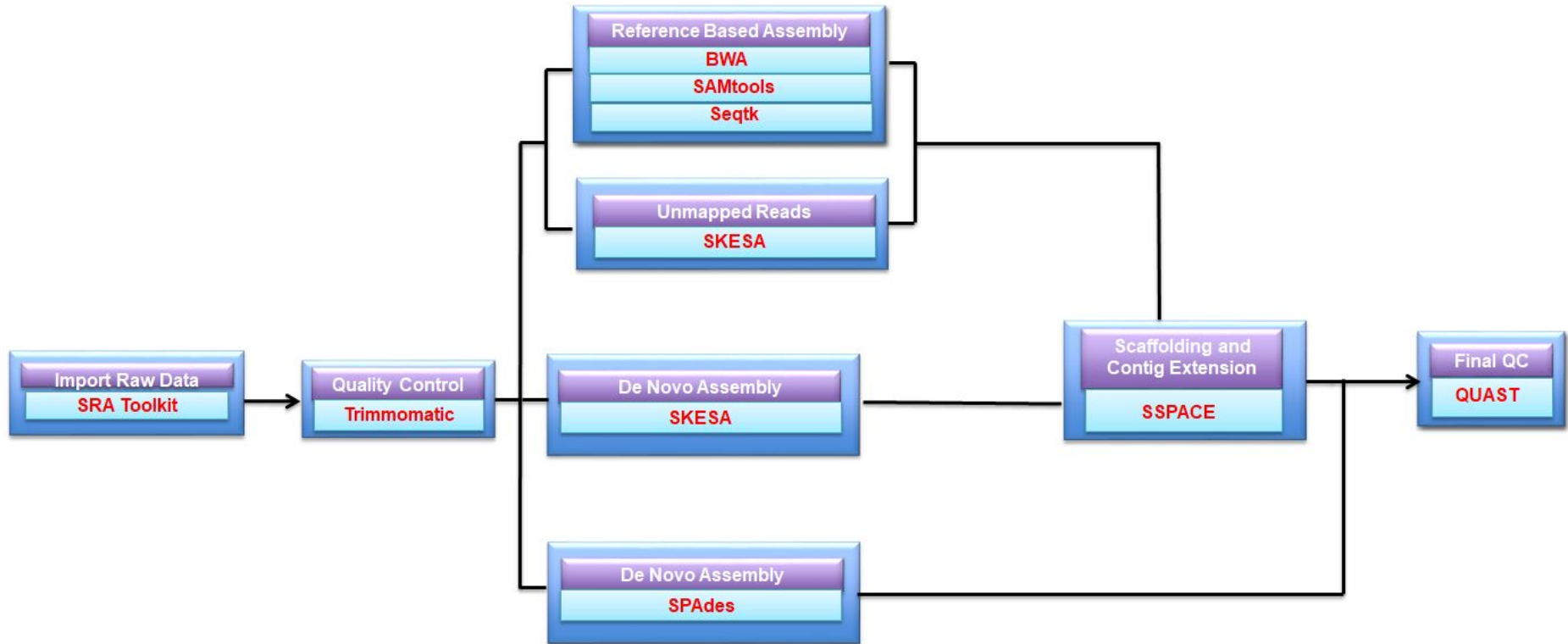
# Final Results, Genome Assembly

BIOL 7210: Computational Genomics - Spring 2018

Team-1 Members: Kunal Agarwal, Victoria Caban, Vasanta  
Chivukula, Seonggeon Cho, Siarhei Hladyshau, Hunter  
Seabolt, Nirav Shah, Tianze Song, Qinwei Zhuang

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# Pipeline

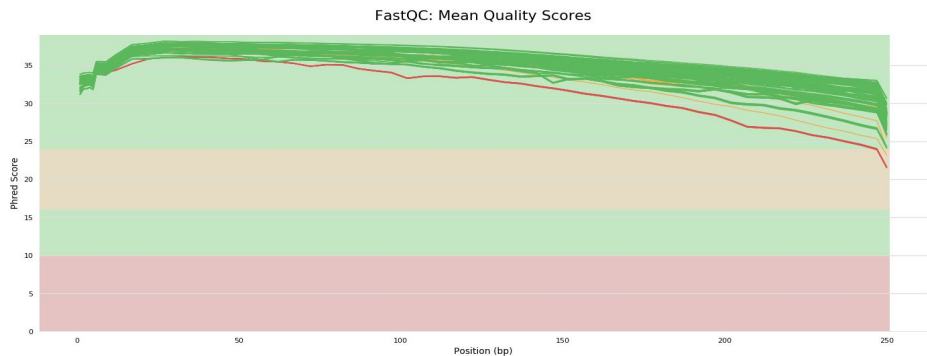


# Trimming and Quality Control

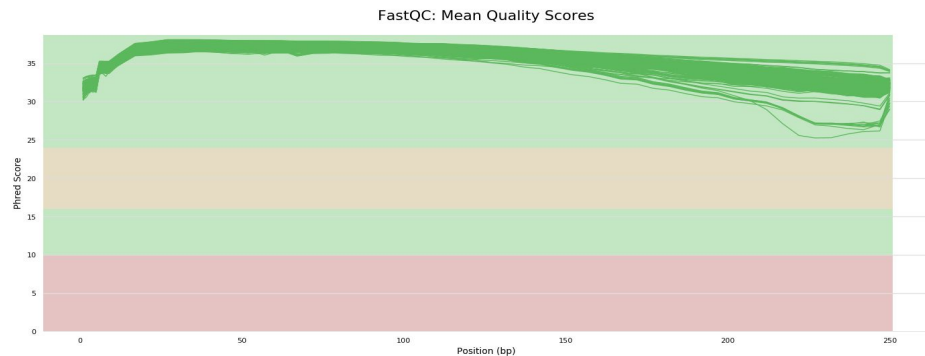
Trimming raw data with Trimmomatic

- ILLUMINACLIP: trims adapter sequences in the reads
- SLIDINGWINDOW: trims the reads based on the threshold quality score set by a user
  - \*4:20 was used in our samples
- MINLEN: drops reads if they are below an assigned length
  - \*20 was set as the minimum length

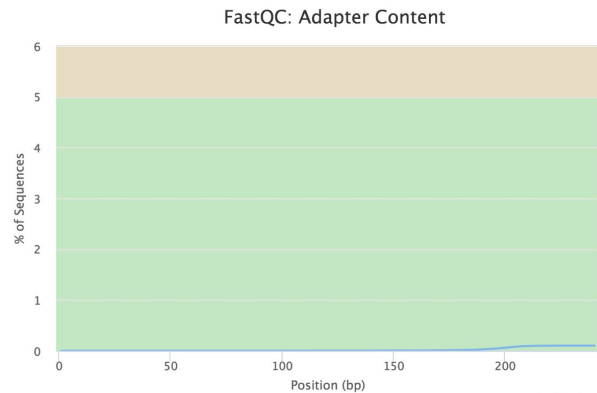
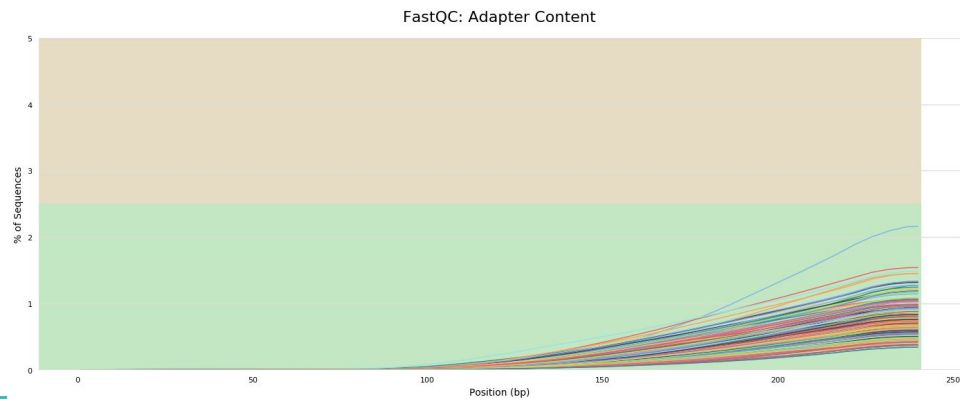
# Trimmomatic Successfully Removes Low Quality and Adapter Reads



Before Trimming

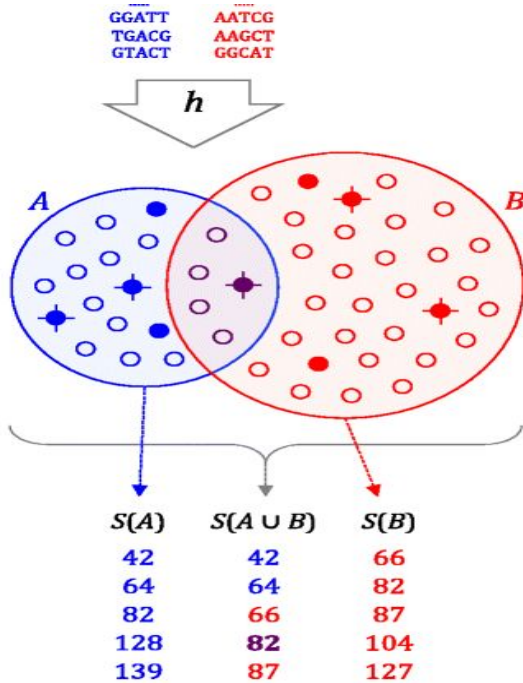


After Trimming



# Reference Based Assembly

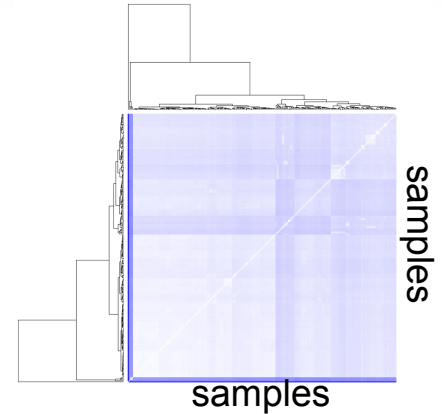
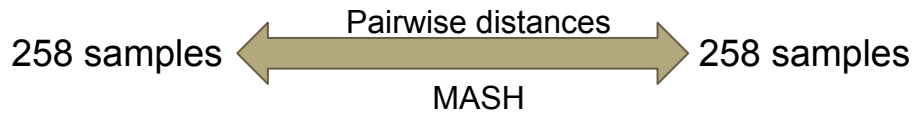
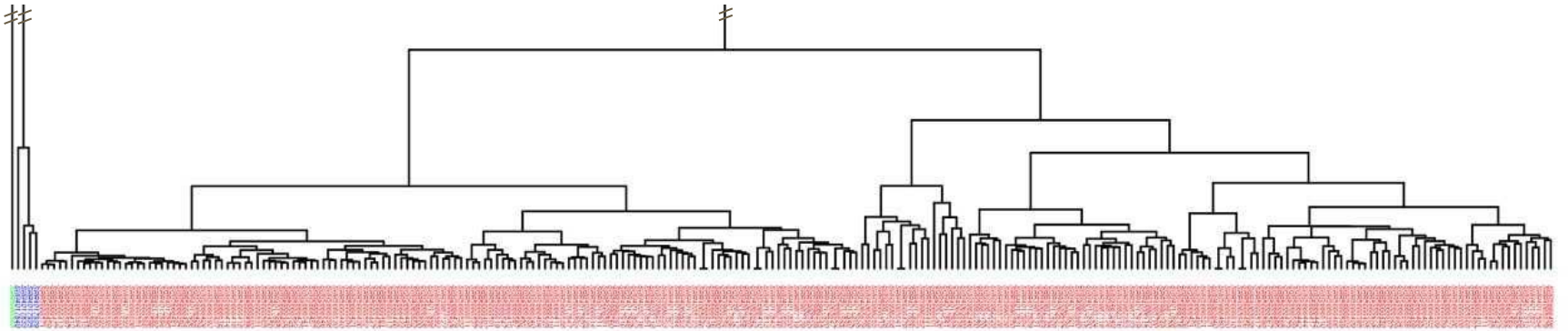
# MASH



- MinHash Algorithm is used by MASH.
- MinHash algorithm provides an estimation of the Jaccard index.
- MASH evaluates mutation distance using Jaccard index between the genomes for similarity.

$$J(A, B) = \frac{|A \cap B|}{|A \cup B|} \approx \frac{|S(A \cup B) \cap S(A) \cap S(B)|}{|S(A \cup B)|}$$

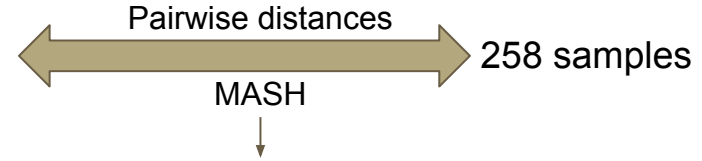
# Evaluation of distance between samples



# Choosing Reference Genomes

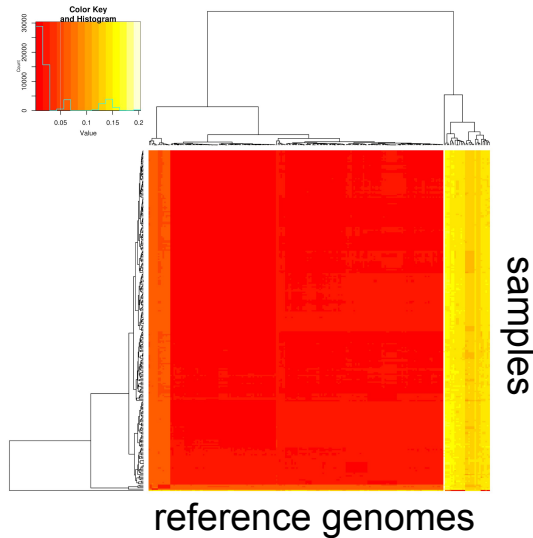


220 complete, reference genomes of *Klebsiella* spp



258 samples

Choice of best genome reference genome for every sample

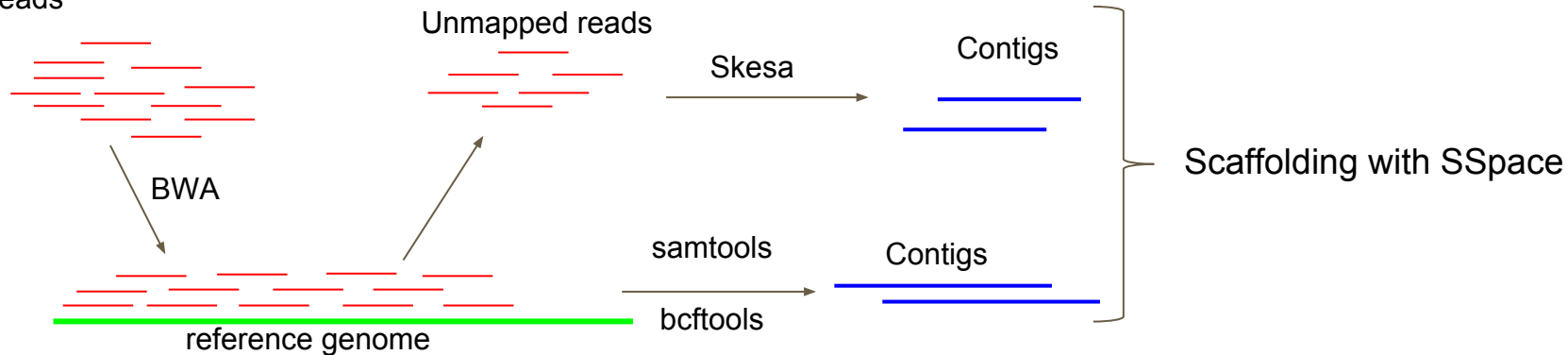


- 252 samples: *Klebsiella pneumoniae*
- 4 samples: *Klebsiella variicola*
- 1 sample: *Klebsiella oxytoca*
- 1 sample: *Klebsiella* sp. 2N3

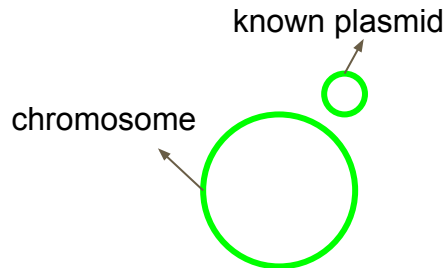


# Reference Based Assembly

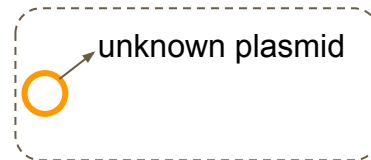
Trimmed reads



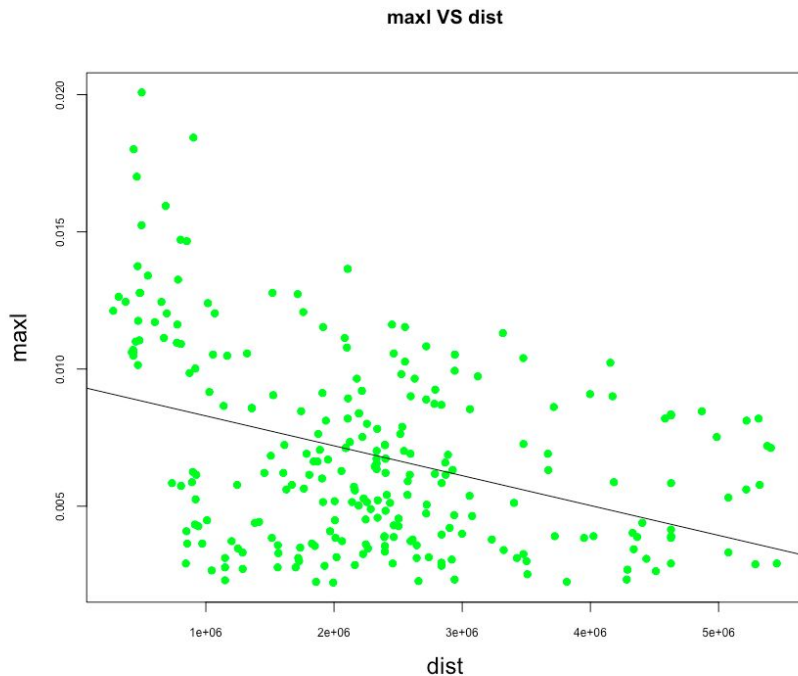
Why to assemble unmapped reads separately?



What if antibiotic resistance is here?



# Importance of reference genome



## Linear Regression

Residuals:

Min	1Q	Median	3Q	Max
-2002720	-836508	-166123	576530	3190016

Coefficients:

	Estimate	Std. Error	t value	Pr(> t )
(Intercept)	3246617	157595	20.601	< 2e-16 ***
all_data\$dist	-137253228	20458750	-6.709	<b>1.25e-10</b> ***

---  
Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*'

Residual standard error: 1146000 on 256 degrees of freedom  
Multiple R-squared: 0.1495, Adjusted R-squared: 0.1462  
F-statistic: 45.01 on 1 and 256 DF, p-value: 1.252e-10

## Pearson's product-moment correlation

t = -6.7088, df = 256, p-value = **1.252e-10**  
alternative hypothesis: true, correlation is not equal to 0  
95 percent confidence interval: -0.4858635, -0.2776702  
sample estimates: cor, -0.3866826

# de Novo Assembly

# de Novo Assembly Using SPAdes



It is an assembler that works based on DeBruijn graphs

Designed to assemble small genome

Do scaffolding by itself

Supports paired-ends and unpaired reads

Give flexibility in Kmer selection

```
Spades.py --careful -k kmer size --pe1-1 forward_paired.fq --pe1-2  
reverse_paired.fq --pe1-s forward_unpaired.fq --pe1-s reverse_unpaired.fq -o  
output_directory
```

# SPAdes Pipeline

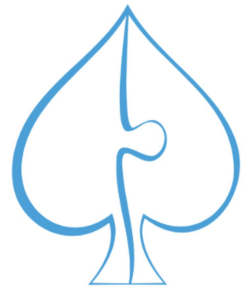


Read Error Correction--BayersHammer

Assemble--Spades

Mismatch Correction--improves mismatch and short indel rates in resulting contigs and scaffolds; this module uses the BWA tool, activated by --careful

# SPAdes Kmer



If we give many kmers in one command line like this:

```
spades.py -k 41,77,99,127 --careful <your reads> -o spades_output
```

Output is the assembly with best N50.

```
spades.py -k 41 --careful <your reads> -o spades_output
```

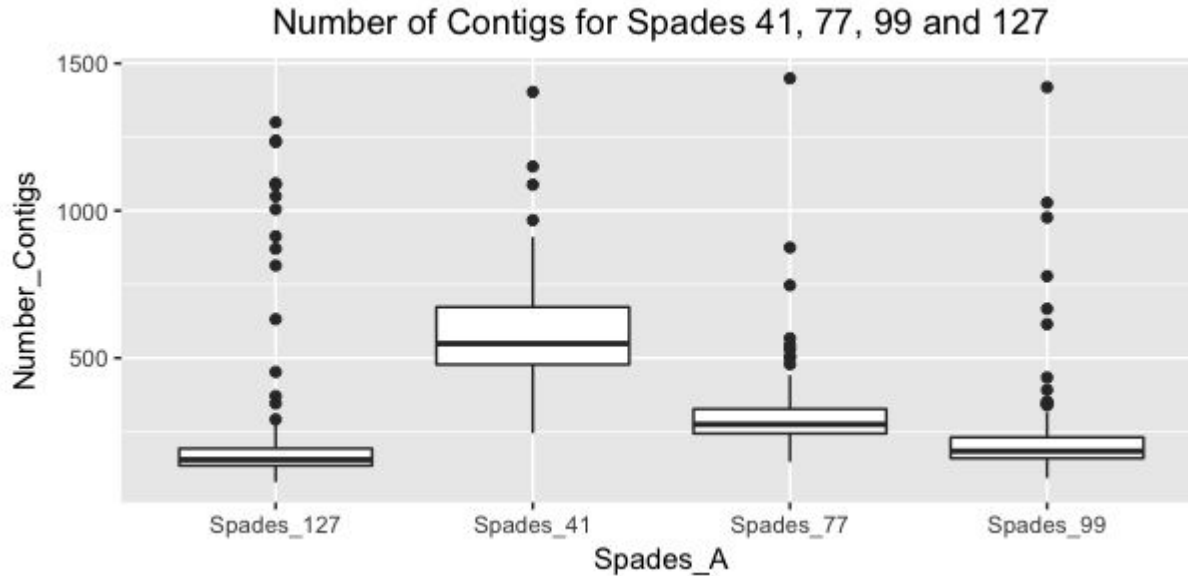
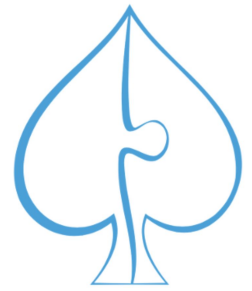
```
spades.py -k 77 --careful <your reads> -o spades_output
```

```
spades.py -k 99 --careful <your reads> -o spades_output
```

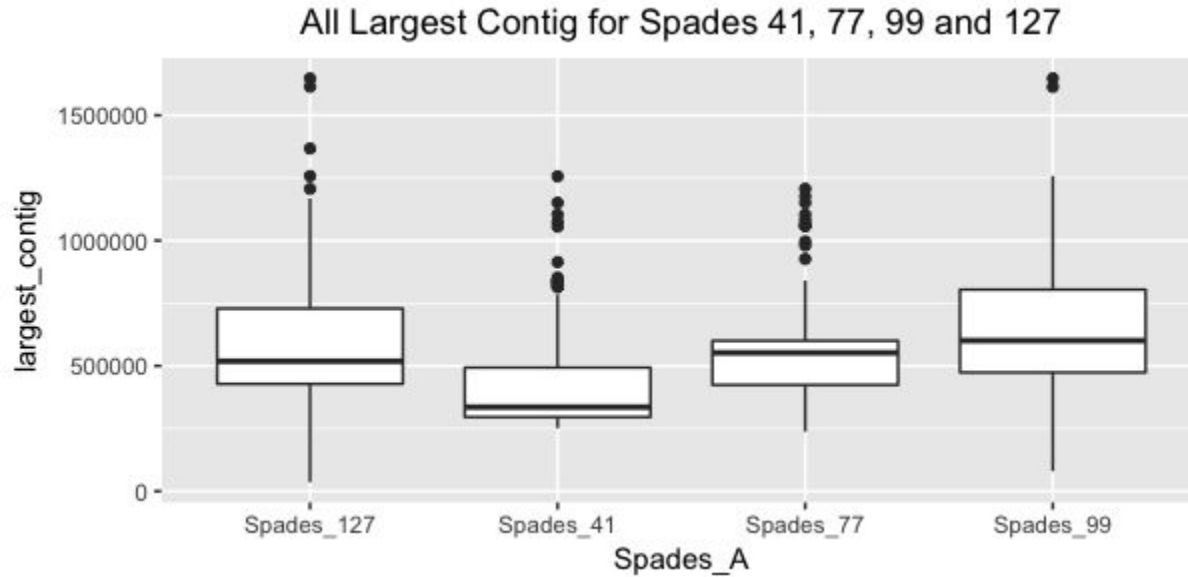
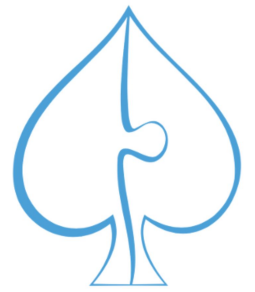
```
spades.py -k 127 --careful <your reads> -o spades_output
```

Select the best assembly by multi-parameters

# SPAdes: number of contigs

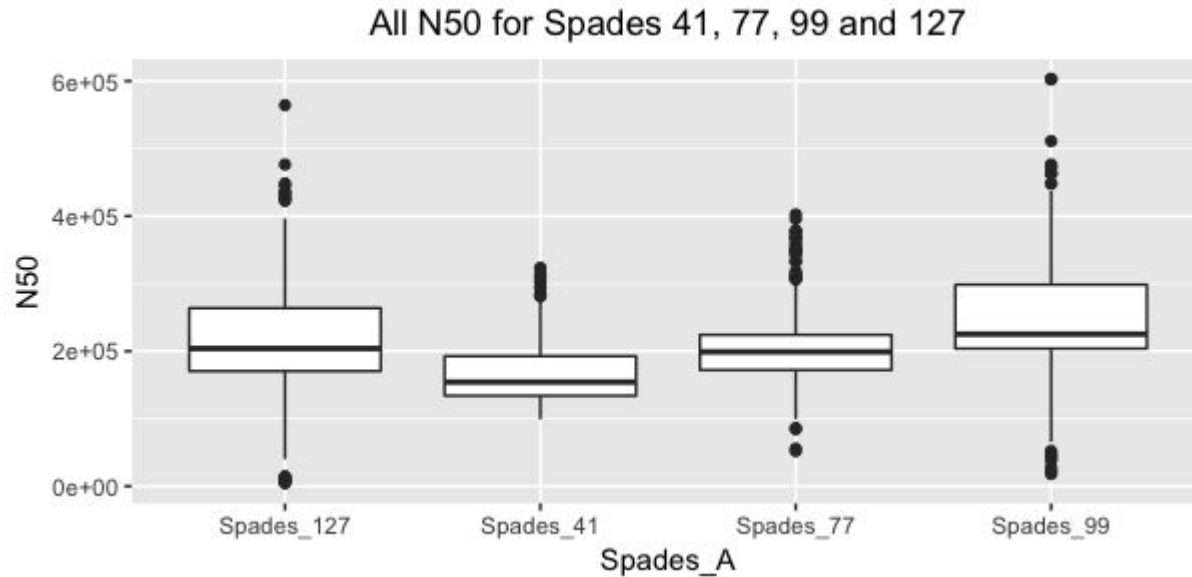
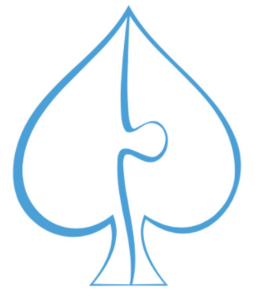


# SPAdes: largest contig





# SPAdes: N50



# de Novo Assembly Using Skesa

- The binary for Skesa was provided by CDC
- It is an assembler that works based on DeBruijn graphs
- It is designed for haploid genomes sequenced using Illumina
- Creates breaks at repeat regions in genomes
- Multi-threaded application - so good for scaling

```
def runSkesa(geneList):
    for a in geneList:
        fFile = '%s_forward_paired.fq' % (a)
        rFile = '%s_reverse_paired.fq' % (a)
        forwardFile = os.path.join(fileDir, fFile)
        reverseFile = os.path.join(fileDir, rFile)
        #print (forwardFile, reverseFile)
        skesaCmd = 'skesa --fastq %s --fastq %s \
--contigs_out /projects/data/team1_genomeAssembly/denovo_skesa/skesaoutput/%s.skesa.fa' % (forwardFile, reverseFile, a)
        os.system(skesaCmd)
```

# Scaffolding Using SSPACE

- Scaffolding Pre-Assemblies After Contig Extension (SSPACE)
- Extends and scaffolds pre-assembled contigs
- Uses Bowtie to map all reads to the pre-assembled contigs
- A library file containing library name, read 1, read 2, insert size (500), error (0.75), FR

```
def generateLibFiles(geneList):
    for gene in geneList:

        libFileName = '%s/%s'%(libFile,gene)

        libText="%s_lib /projects/data/team1_genomeAssembly/trimming2/fastq/trimmed/%s_forward_paired.fq \
/projects/data/team1_genomeAssembly/trimming2/fastq/trimmed/%s_reverse_paired.fq 250 0.75 FR" %(gene,gene,gene)
        if not os.path.exists(libFileName):
            with open(libFileName,'w') as fh:
                fh.write(libText)
            fh.close()
```

SSPACE basic

FOR 100% BASECLEAR

# Scaffolding continued

- Contig extension was performed using SSAKE method by changing the standard -x 0 to 1
- This is followed by building scaffolds and merging contigs
- The output contains final scaffolds in fasta format, scaffolds with initial numbered contigs, a log file and a summary file

Running the SSPACE command for scaffolding using default parameters and contig extension (-x 1)

```
sspaceCmd = "perl /projects/data/team1_genomeAssembly/SSPACE/sspace_basic/SSPACE_Basic.pl -l \  
/projects/data/team1_genomeAssembly/denovo_skesa/sspaceLibrary/%s \  
-s /projects/data/team1_genomeAssembly/denovo_skesa/skesaoutput/%s.skesa.fa \  
-x 1 -T 8 -b %s.sspace -m 20 -o 15 -a 0.8 -n 12 -g 3 -p 1" %(gene,gene,gene)
```

```
os.system(sspaceCmd)  
print("Done scaffolding")
```



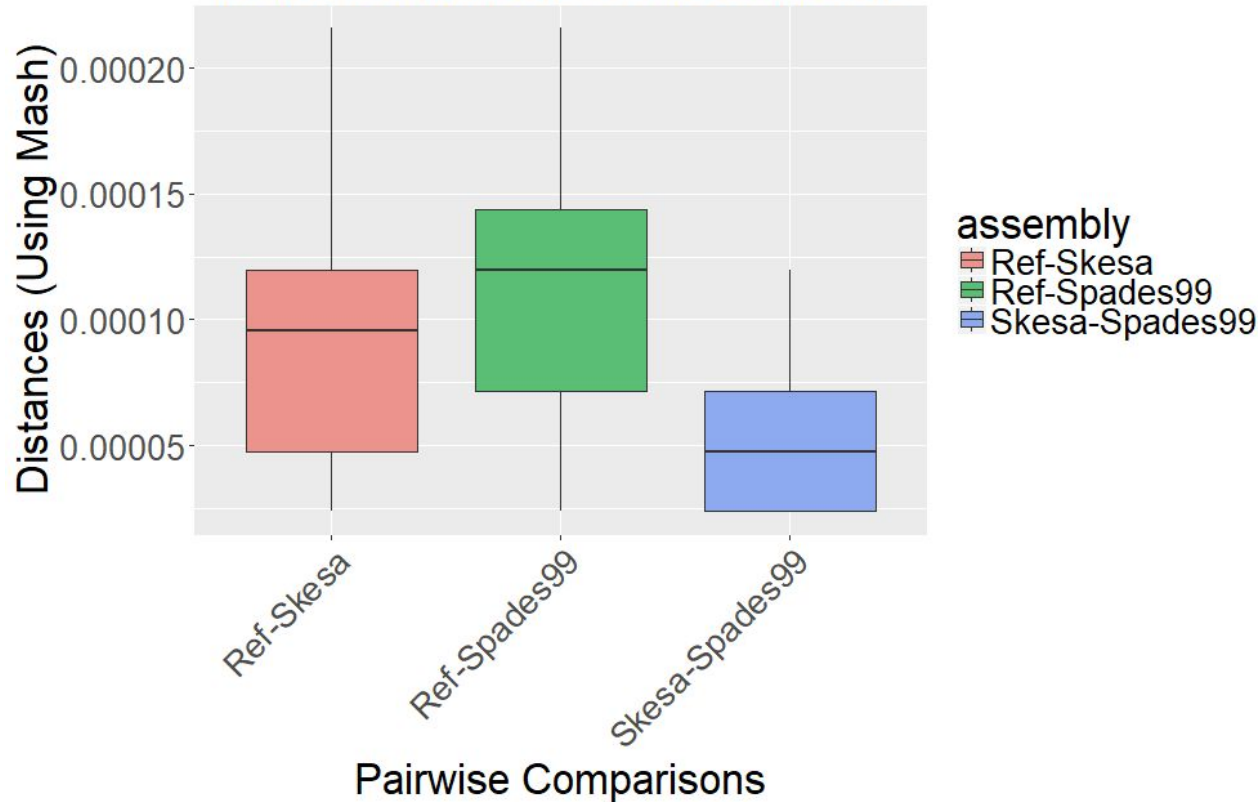
# Comparison between Spades and Skesa

Parameters	Average SPAdes	Average Skesa	P value
N50	250137	229259	0.19592
# Contigs	212	123	1.55E-10***
Largest Contigs	645324	609123	0.063028
Total Length	5588948	5601627	0.44905
N's per 100kbp	2.781	11.456	0.000104***

# Merging assemblies



## Mash Distances of Assemblies

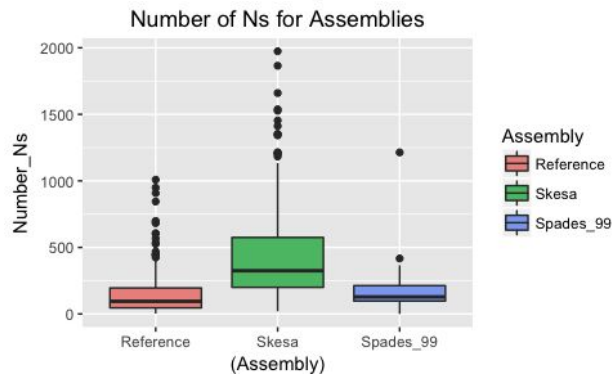
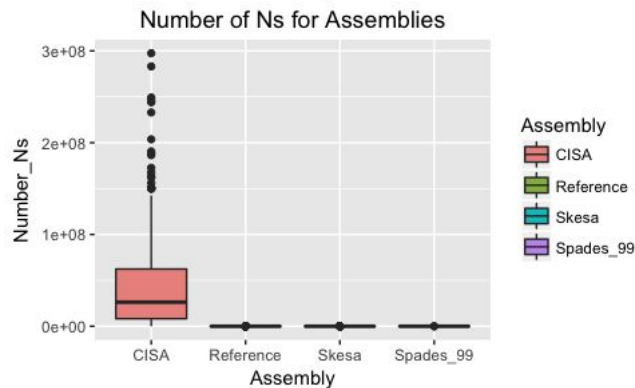
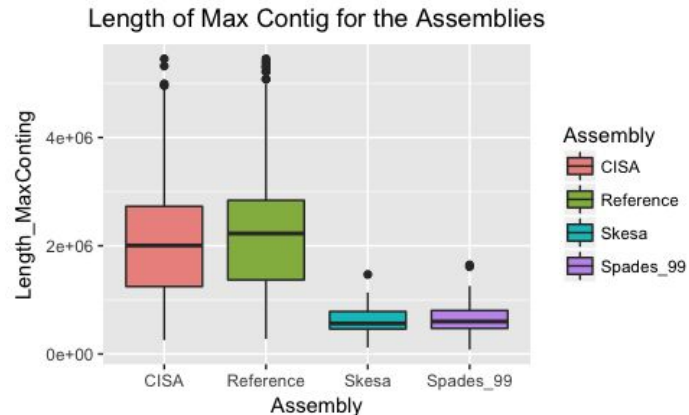
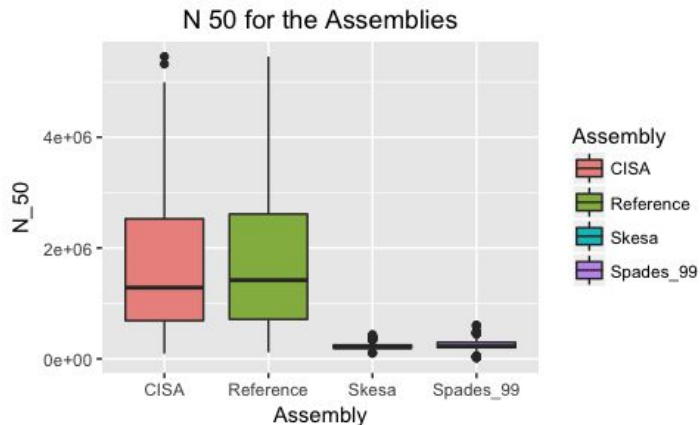


Reference based

Skesa

CISA

# Quality of assemblies



# References

Bankevich, Anton et al. “SPAdes: A New Genome Assembly Algorithm and Its Applications to Single-Cell Sequencing.” *Journal of Computational Biology* 19.5 (2012): 455–477. *PMC*. Web. 6 Mar. 2018.

Bolger, Anthony M., Marc Lohse, and Bjoern Usadel. “Trimmomatic: A Flexible Trimmer for Illumina Sequence Data.” *Bioinformatics* 30.15 (2014): 2114–2120. *PMC*. Web. 6 Mar. 2018.

Gurevich, Alexey et al. “QUAST: Quality Assessment Tool for Genome Assemblies.” *Bioinformatics* 29.8 (2013): 1072–1075. *PMC*. Web. 6 Mar. 2018

Heng Li, Richard Durbin; Fast and accurate short read alignment with Burrows–Wheeler transform, *Bioinformatics*, Volume 25, Issue 14, 15 July 2009, Pages 1754–1760, <https://doi.org/10.1093/bioinformatics/btp324>

Boetzer M, Henkel CV, Jansen HJ, Butler D and Pirovano W. 2010. Scaffolding pre-assembled contigs using SSPACE. *Bioinformatics*. 27(4):578-579

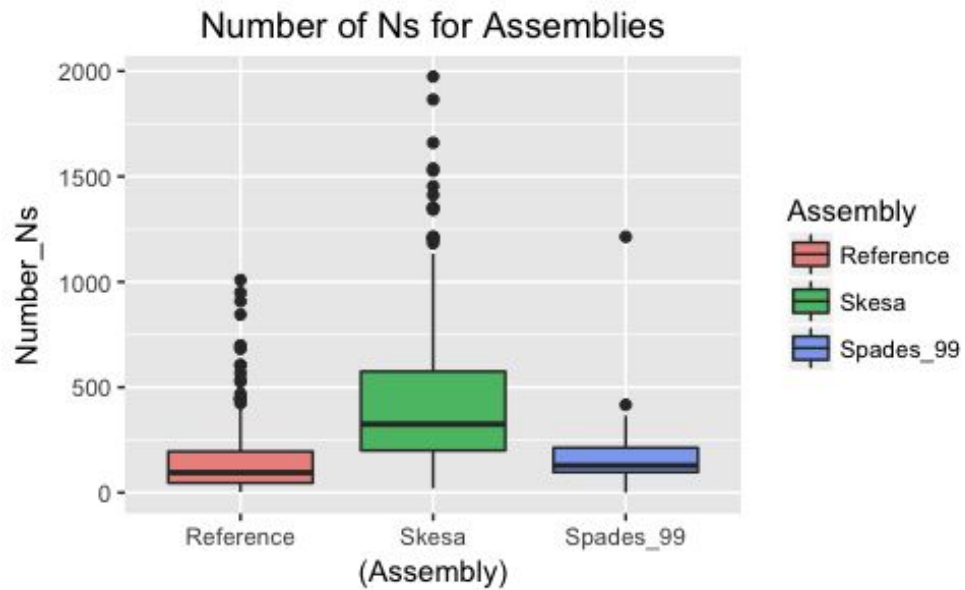
<http://bioinf.spbau.ru/spades>

<http://sb.nhri.org.tw/CISA/en/Instruction>

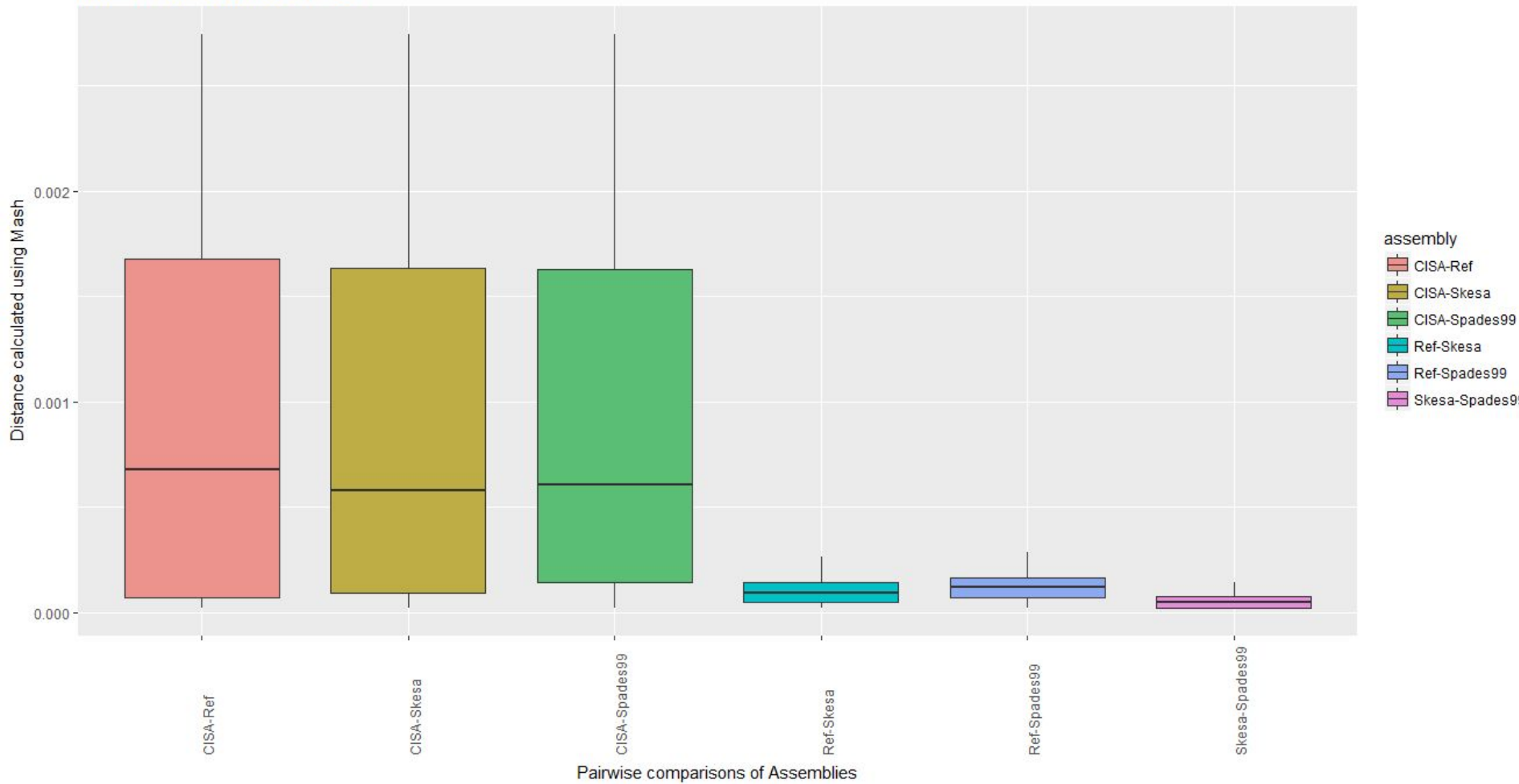
<https://github.com/enormandea>



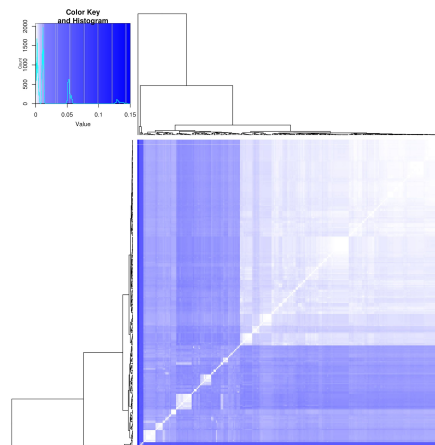
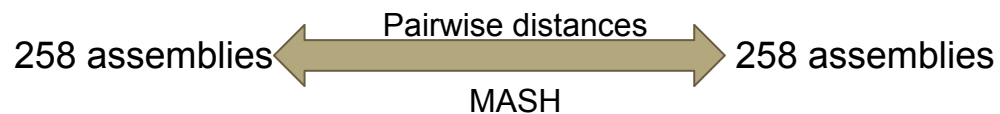
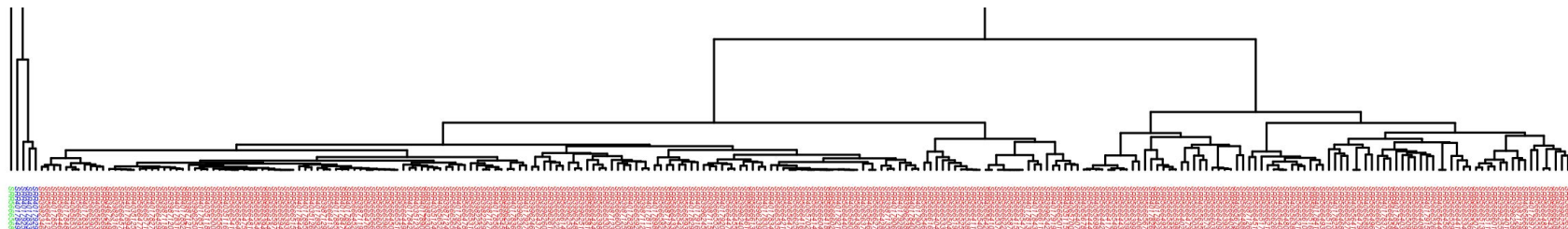
**Thank you for your attention!**



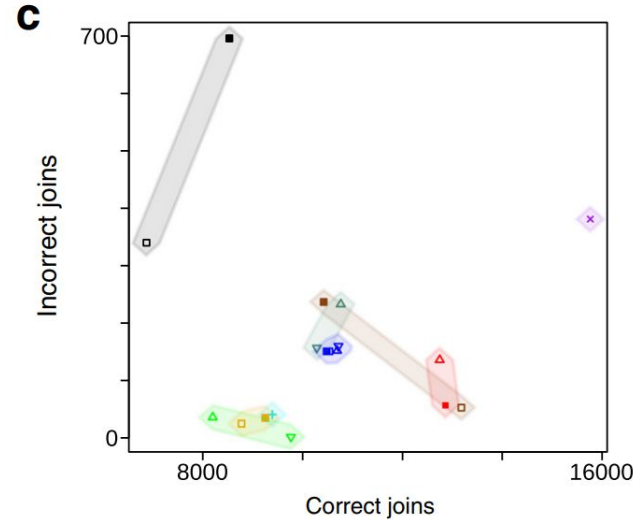
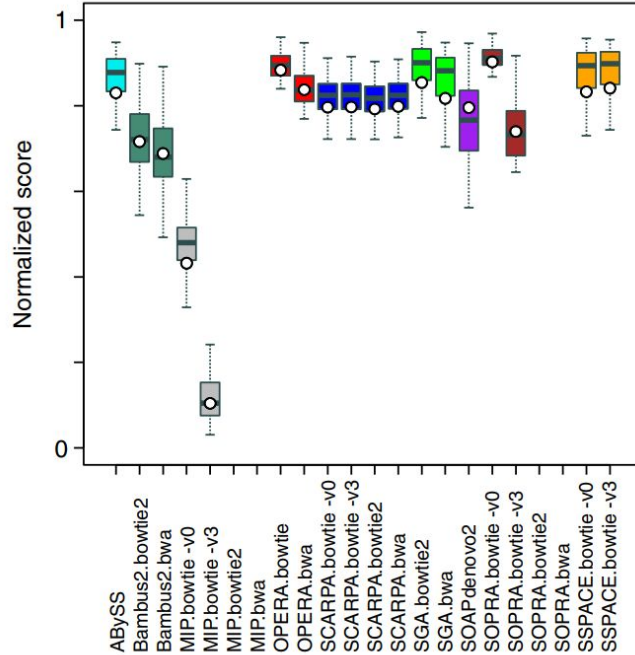
Mash Distances of Assemblies



# Evaluation of assemblies

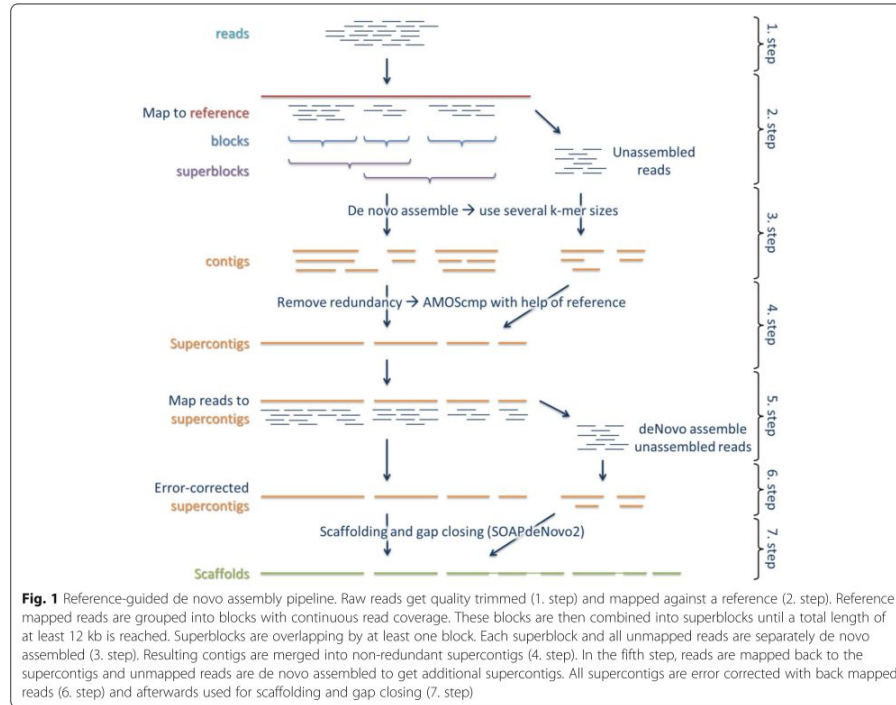


# Choice of scaffolding tool



Genome Biol. 2014; 15(3),Martin Hunt, et.al.

# Example of pipeline for reference guided assembly



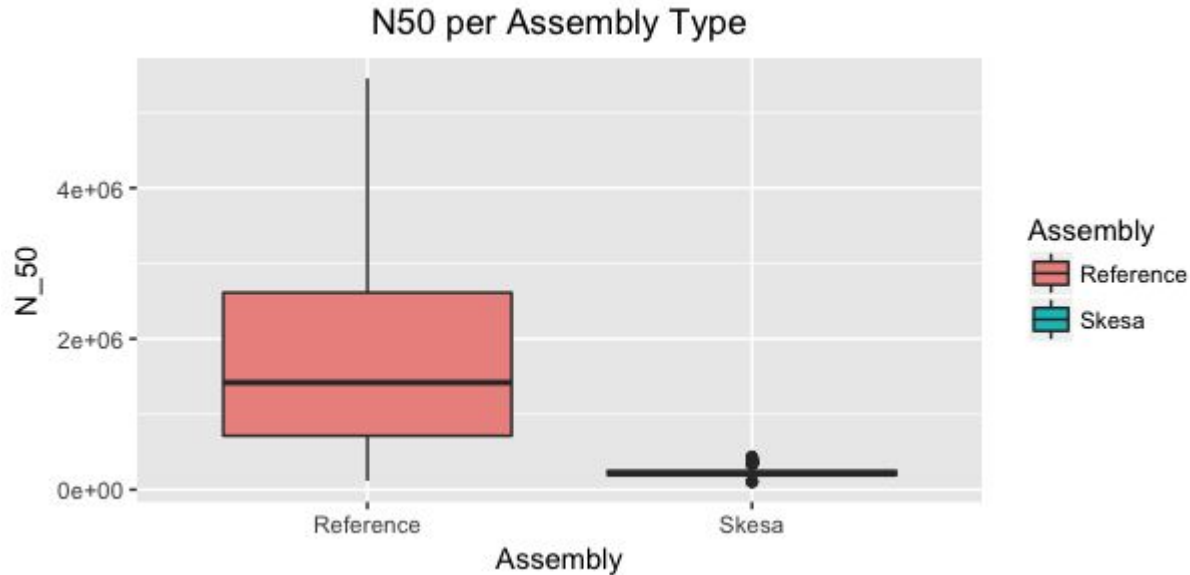
BMC Bioinformatics. 2017 Nov 10;18(1):474.

Lischer HEL, Shimizu KK.

# Pipeline for reference based assembly

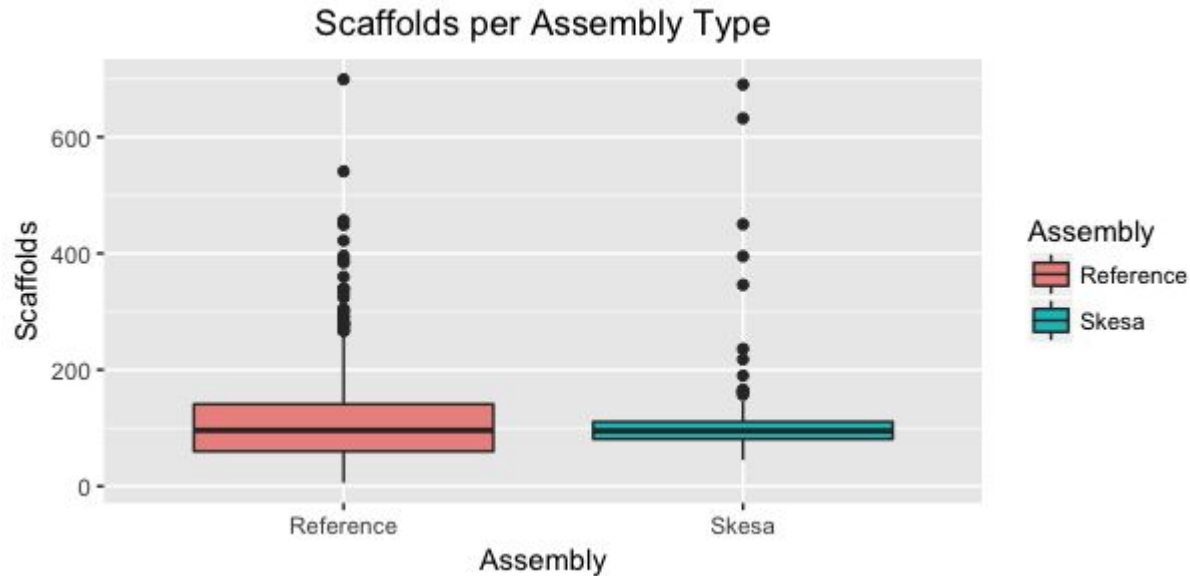
```
bwa index -a is [reference genome]
bwa mem [reference genome] [forward and reverse reads] > [output.sam]
samtools sort [output.sam] > [output_sorted.bam]
samtools index [output_sorted.bam]
samtools view -b -f 4 [output_sorted.bam] > [unmapped.bam]
samtools bam2fq [unmapped.bam] > [unmapped.fastq]
samtools mpileup -v --no-BAQ -f [reference genome] [output_sorted.bam] |
  bcftools call -c | vcfutils.pl vcf2fq | seqtk seq -A > [assembly.fasta]
```

# N50 for Referenced Based and Skesa Assembly

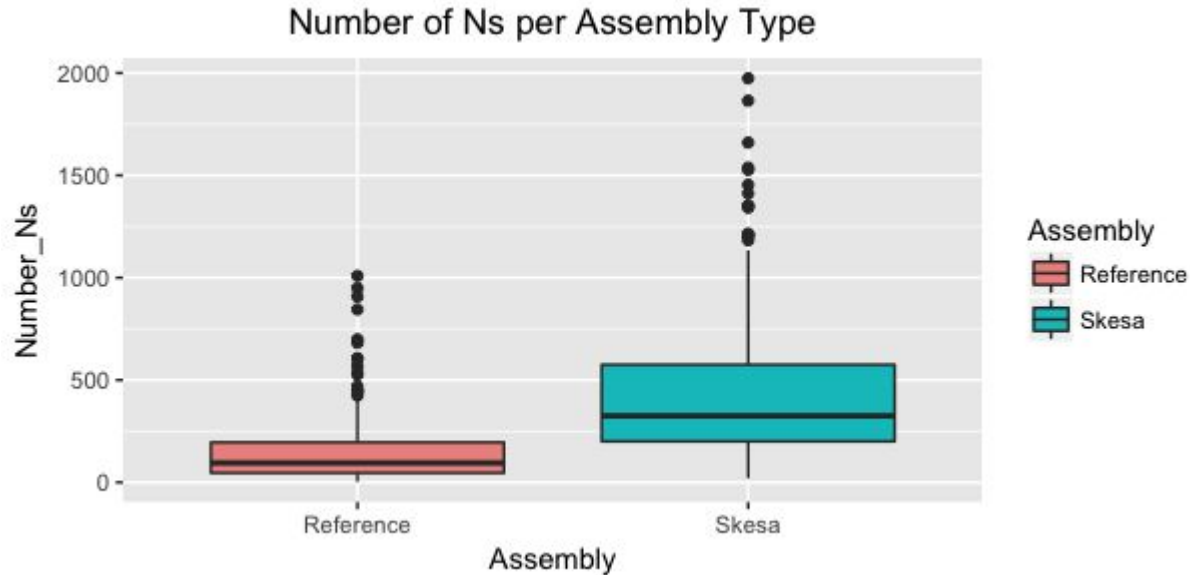




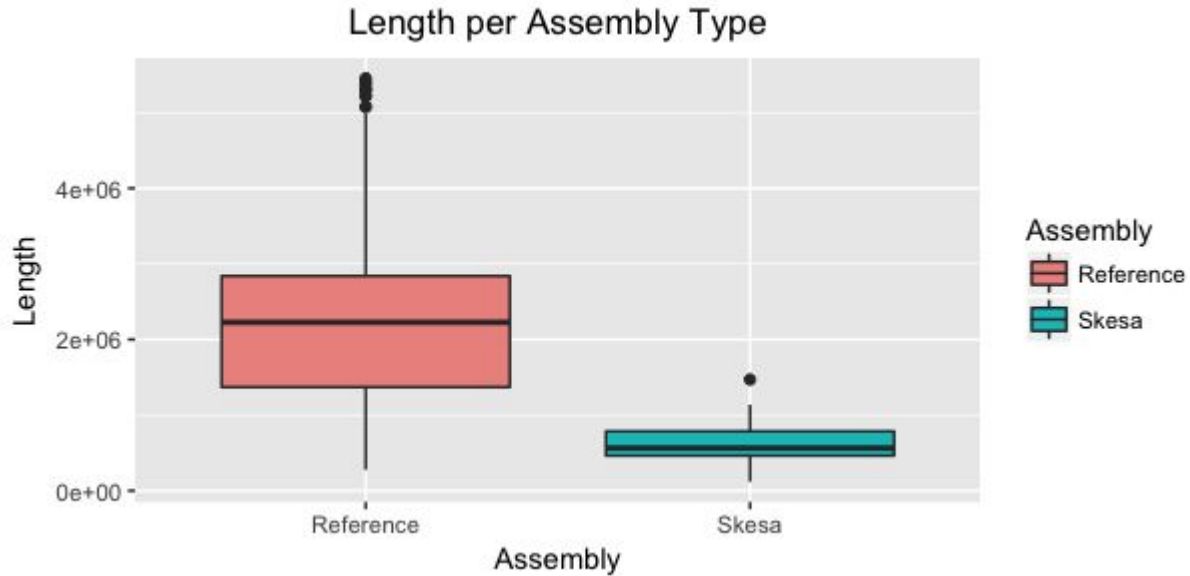
# Scaffolds for Referenced Based and Skesa Assembly



# Number of Ns for Referenced Based and Skesa Assembly



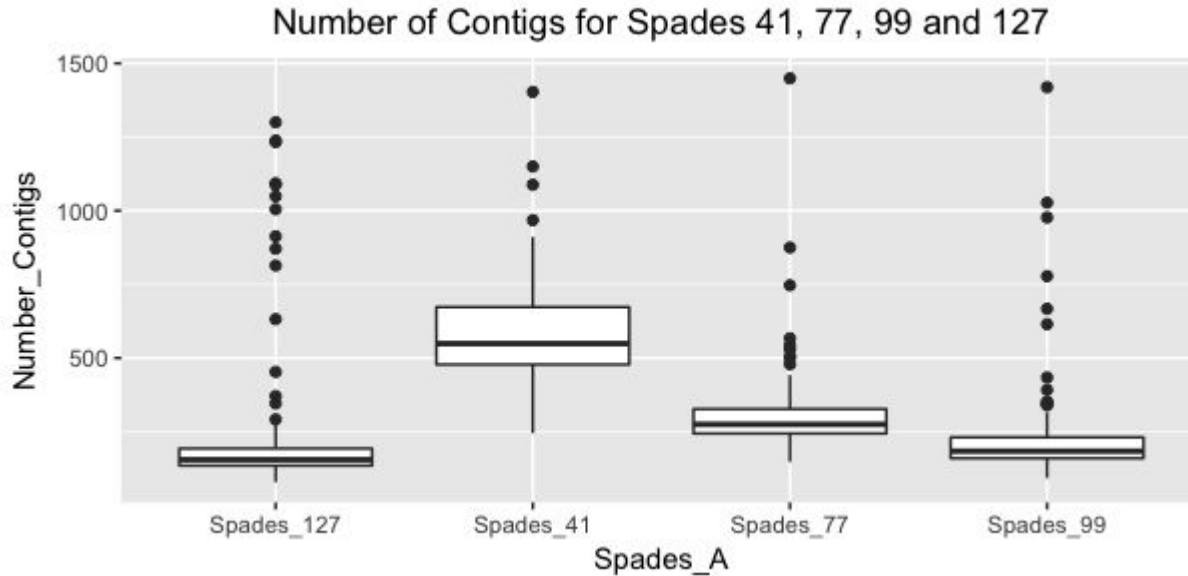
# Length for Referenced Based and Skesa Assembly



# References

<https://doi.org/10.1186/s13059-016-0997-x>

# Number of Contigs for Spades Assembly



Parameter

Table Analyzed **spades\_number\_of\_contigs**

### One-way analysis of variance

P value < 0.0001  
P value summary \*\*\*  
Are means signif. different? (P < 0.05) Yes  
Number of groups 4  
F 146.7  
R square 0.3006

Bartlett's test for equal variances

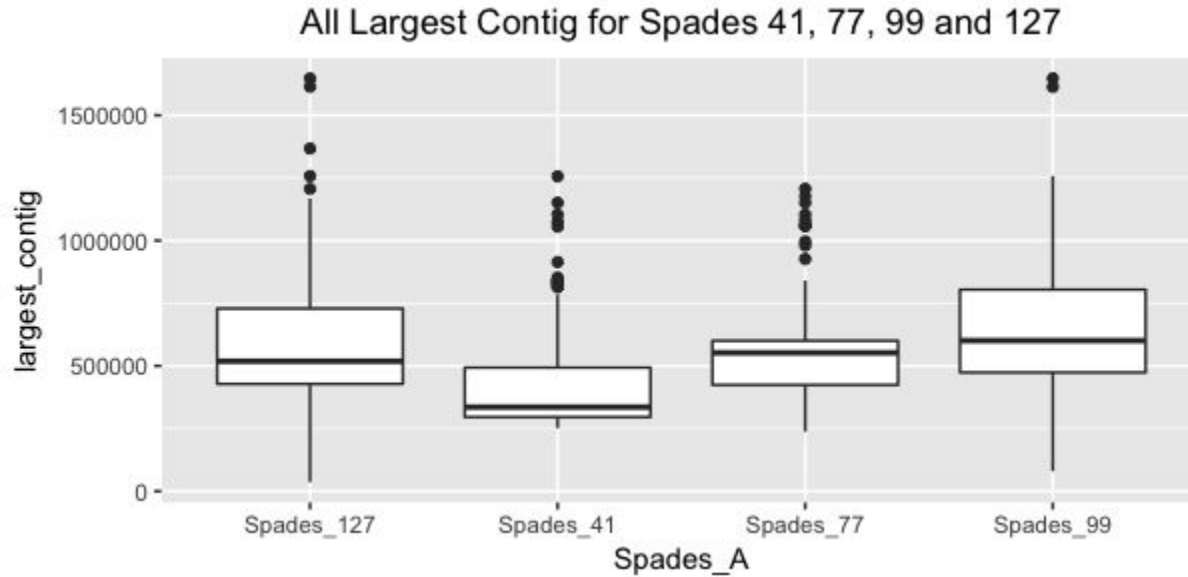
Bartlett's statistic (corrected) 28.16  
P value < 0.0001  
P value summary \*\*\*  
Do the variances differ signif. (P < 0.05) Yes

ANOVA Table	SS	df	MS
Treatment (between columns)	2.524e+007	3	8.414e+006
Residual (within columns)	5.872e+007	1024	57343
Total	8.396e+007	1027	

Tukey's Multiple Comparison Test	Mean Diff.	q	Significant? P < 0.05?	95% CI of diff
SPAdes 41 vs SPAdes 77	292.1	19.56	Yes ***	237.3 to 346.9
SPAdes 41 vs SPAdes 99	379.2	25.38	Yes ***	324.4 to 434.0
SPAdes 41 vs SPAdes 127	384.3	25.72	Yes ***	329.5 to 439.1
SPAdes 77 vs SPAdes 99	87.03	5.826	Yes ***	32.23 to 141.8
SPAdes 77 vs SPAdes 127	92.12	6.167	Yes ***	37.32 to 146.9
SPAdes 99 vs SPAdes 127	5.089	0.3407	No ns	-49.71 to 59.89

**Summary: SPAdes 99 and SPAdes 127 have the significantly lower contig number compared to other kmer size**

# Largest Contig for Spades Assembly



Parameter

Table Analyzed **spades\_large\_contigs**

# Summary: SPAdes 99 has the significantly longer contig length compared to other kmer size

## One-way analysis of variance

P value < 0.0001  
P value summary \*\*\*  
Are means signif. different? (P < 0.05) Yes  
Number of groups 4  
F 47.50  
R square 0.1216

## Bartlett's test for equal variances

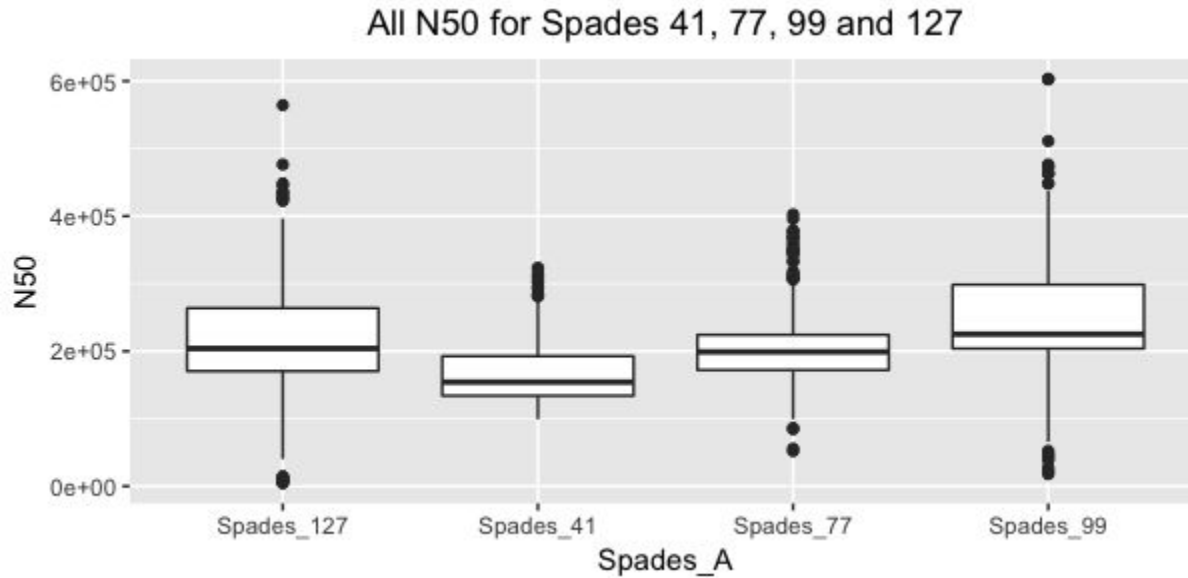
Bartlett's statistic (corrected) 52.39  
P value < 0.0001  
P value summary \*\*\*  
Do the variances differ signif. (P < 0.05) Yes

ANOVA Table	SS	df	MS
Treatment (between columns)	6.592e+012	3	2.197e+012
Residual (within columns)	4.760e+013	1029	4.626e+010
Total	5.419e+013	1032	

Tukey's Multiple Comparison Test	Mean Diff.	q	Significant? P < 0.05?	95% CI of diff
SPAdes 41 vs SPAdes 77	-124376	9.280	Yes ***	-173547 to -75205
SPAdes 41 vs SPAdes 99	-220155	16.47	Yes ***	-269185 to -171126
SPAdes 41 vs SPAdes 127	-153780	11.46	Yes ***	-202999 to -104562
SPAdes 77 vs SPAdes 99	-95779	7.174	Yes ***	-144761 to -46797
SPAdes 77 vs SPAdes 127	-29404	2.194	No ns	-78575 to 19767
SPAdes 99 vs SPAdes 127	66375	4.966	Yes **	17345 to 115405



# N50 for Spades Assembly



Parameter  
Table Analyzed **N50**

## Summary: SPAdes 99 has the significantly higher N50 compared to other kmer size

### One-way analysis of variance

P value < 0.0001  
P value summary \*\*\*  
Are means signif. different? (P < 0.05) Yes  
Number of groups 4  
F 44.23  
R square 0.1147

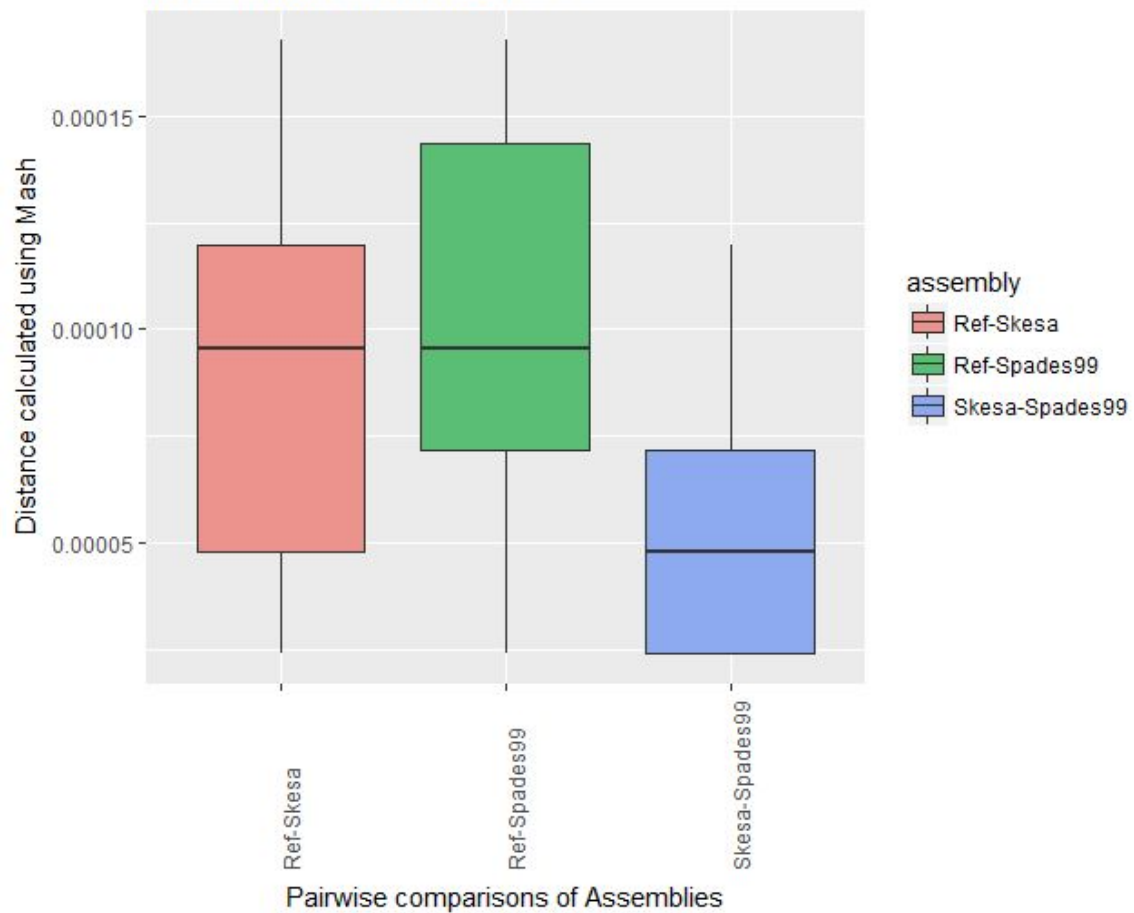
### Bartlett's test for equal variances

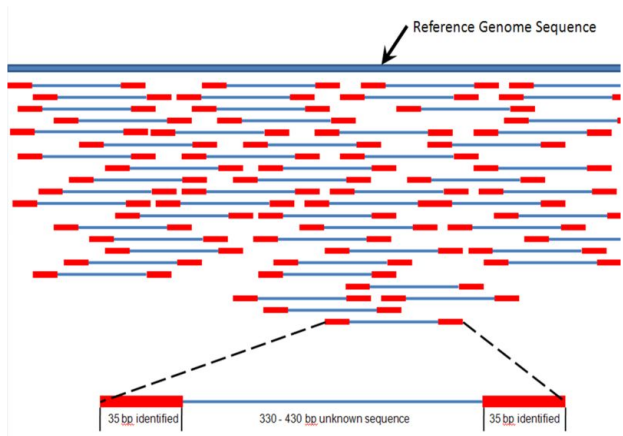
Bartlett's statistic (corrected) 143.6  
P value < 0.0001  
P value summary \*\*\*  
Do the variances differ signif. (P < 0.05) Yes

ANOVA Table	SS	df	MS
Treatment (between columns)	7.819e+011	3	2.606e+011
Residual (within columns)	6.034e+012	1024	5.893e+009
Total	6.816e+012	1027	

Tukey's Multiple Comparison Test	Mean Diff.	q	Significant? P < 0.05?	95% CI of diff
SPAdes 41 vs SPAdes 77	-36846	7.695	Yes ***	-54414 to -19279
SPAdes 41 vs SPAdes 99	-77679	16.22	Yes ***	-95247 to -60111
SPAdes 41 vs SPAdes 127	-43688	9.124	Yes ***	-61256 to -26121
SPAdes 77 vs SPAdes 99	-40833	8.527	Yes ***	-58401 to -23265
SPAdes 77 vs SPAdes 127	-6842	1.429	No ns	-24410 to 10725
SPAdes 99 vs SPAdes 127	33991	7.098	Yes ***	16423 to 51558

## Mash Distances of Assemblies





```

1      11     21     31     41     51     61     71     81
AGCTTTTCATTCTGACTGCAACGGGCAATATGTCTCTGTGTGGATTAAAAAAGAGTGTCTGATAGCAGCTTCTGAAGCTGGTACCTGCGGTGAGTA
|.....
.....C.....
.....C.....G.....G.....A.....
.....G.....C.....C.....G.....
.....T.....A.....T.....
.....C.....G.....
.....C.....A.....A.....A.....
.....C.....G.....A.....A.....G.....
.....T.....C.....A.....
.....C.....T.....C.....G.....
.....T.....A.....A.....
.....T.....A.....G.....
.....T.....T.....T.....
.....T.....

```

# Supplementary: SPAdes Kmer Selection

For multicell paired end 250bp data:

It suggests:

```
spades.py -k 21,33,55,77,99,127 --careful <your reads> -o spades_output
```

Kmer selection can be tricky.